

# IMPORTANCE OF OXYGEN TENSION IN HUMAN OVARIAN TISSUE IN VITRO CULTURE

**Francisco Vitale**, Luciana Cacciottola, Fang Shu Yu, Marta Barretta, Camille Hossay, Jacques Donnez, Marie-Madeleine Dolmans.

*Gynecology Research Unit, Institut de Recherche Expérimentale et Clinique, Université Catholique de Louvain, Brussels, Belgium, Pôle de Recherche en Gynécologie, Institut de Recherche Expérimentale et Clinique, Université Catholique de Louvain, Brussels, Belgium, Gynecology Research Unit, Institut de Recherche Expérimentale et Clinique, Université Catholique de Louvain, Brussels, Belgium, Department of Biomedical Science for the Health, University of Milan, Milan, Italy, Gynecology Research Unit, Institut de Recherche Expérimentale et Clinique, Université Catholique de Louvain, Brussels, Belgium, Society for Research into Infertility, Brussels, Belgium Professor Emeritus at Université Catholique de Louvain, Brussels, Belgium, Pôle de Recherche en Gynécologie, Institut de Recherche Expérimentale et Clinique, Université Catholique de Louvain, Brussels, Belgium Gynecology Department, Cliniques Universitaires Saint-Luc, Brussels, Belgium.*

## Abstract Body

**Background:** The primordial follicle (PMF) pool resides within the ovarian cortex, where oxygen tension is 2-8% O<sub>2</sub>. However, in vitro culture (IVC) is commonly performed at atmospheric oxygen tension (20% O<sub>2</sub>), and still shows unfavorable results.

**Objective:** To improve the viability of the PMF pool by modulating IVC oxygen tension.

**Materials and methods:** Frozen-thawed cortical fragments from 6 women were cultured for 6 days at (i) 20% O<sub>2</sub>, (ii) 5% O<sub>2</sub>, and (iii) using a hypoxic preconditioning (HPC) protocol (24 hours at 1% O<sub>2</sub>, followed by 5 days at 5% O<sub>2</sub>). Non-cultured fragments served as controls. Analyses were conducted based on follicle classification (H&E staining), follicle proliferation (Ki67 immunolabeling), follicle apoptosis (caspase-3 immunostaining), oxidative stress injury (8-hydroxy-2-deoxyguanosine immunofluorescence), DNA damage (phosphorylated histone H2AX immunofluorescence) and cellular senescence (β-galactosidase staining). Glucose and lactate medium levels were evaluated every two days for tissue metabolic activity analysis.

**Results:** The 20% and 5% O<sub>2</sub> groups showed significantly ( $p < 0.0001$ ) decreased PMF proportions compared to the control group, but 5% O<sub>2</sub> yielded significantly ( $p = 0.0006$ ) higher PMF rates than 20% O<sub>2</sub>. Percentages of apoptotic follicles ( $p = 0.003$ ), oxidative stress-associated lesions ( $p < 0.0001$ ), DNA damage ( $p = 0.02$ ), and cellular senescence ( $p < 0.001$ ) were significantly lower in the 5% O<sub>2</sub> group than the 20% O<sub>2</sub> group. Moreover, PMFs exposed to 20% O<sub>2</sub> conditions exhibited a significantly ( $p = 0.01$ ) superior proliferation rate compared to 5% O<sub>2</sub>.

Apoptosis rates were 2-fold higher in the HPC group than the 5% O<sub>2</sub> group ( $p = 0.0004$ ), probably due to insufficient oxygen supply. The former also displayed massive lactate release to the extracellular compartment.

**Conclusion:** IVC at 5% O<sub>2</sub> resulted in better follicle viability than did 20% O<sub>2</sub> or HPC conditions, mainly by protecting the PMF pool against oxidative injury and ensuing DNA damage.